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Dip/Contact-Slides for testing the number of bacteria in liquids and on surfaces



Before use, allow the Dip-Slides to reach room temperature for 5 minutes (do not open the tube!). Remove the Dip-Slide from the tube for use, avoid contact of the agar surface with the skin, do not breathe onto the test sample or sneeze!

Immersion test specimens for water (not drinking water), water-based dispersions, emulsions (cooling lubricants), paints and other aqueous solutions.

Contact plates for surface testing: Place the Dip-Slide at a slight angle on a solid surface and press down slowly until the bending angle between the cap and the culture medium is approx. 90° . Press on the surface for ~10 seconds and roll from side to side gently. Repeat the process with the second side at nearby location.

Light yellow TSA-Medium with disinfectant neutralizer for aerobic total plate count, most colonies appear as red dots. Colorless or faintly colored dots should be counted as well.

Light pink Rose Bengal-Medium for fungi. Colonies appear as fibers, and yeasts appear as light pink dots. Dichloran, rose bengal, gentamicin and trimethoprim inhibit bacterial growth, and reduce the colony sizes of rapidly growing molds. Rose Bengal may stain colonies pink.

Storage conditions and duration of use

Store at 10-25°C, protected from direct sunlight. Expiry date of the unopened Dip-Slide is stated on the product label. Don't use the product after the expiry date! If the unopened product shows signs of contamination - dump it. Opened products must be used immediately.

Warnings and precautions

The tests must be carried out by trained personnel. The test method is part of the instructions for use. Modifications and other applications must be validated by the user on his own responsibility.

Contact with skin or breath will contaminate culture medium carriers and make them unusable, please work cleanly!

In case of accidental contact with incubated Dip-Slides, clean the skin immediately with a suitable skin disinfectant. (Generally, thorough washing with soap should be sufficient).

During incubation, the tube lid may only be screwed on loosely (air supply and avoidance of condensation water).

For postal and messenger shipping, the lid must be tightly closed (screwed shut)!

Dip-Slides or germ testers should only be used once.

Disposal

Seal used tubes and take them to an incineration plant for household waste in a tightly closed container/tear-proof bag. Do not open used tubes!

Dip/Contact-Slides for testing the number of bacteria in liquids and on surfaces



User manual:

- 1. Open the screw cap of the tube, dip the Dip-Slide into the sample, both sides must be completely wetted. If only small amounts of sample are present, pour the sample over both sides of the culture medium as evenly as possible.
- 2. Drain off excess sample and wipe off the Dip-Slide at the lower end. Do not touch the culture medium, especially not with your fingers.
- 3. Put the Dip-Slide back into the tube and screw it down loosely.
- 4. Fill in the label and paste it on the tube.

5 Incubate the Dip-Slides at approx. 25°C for 3-5 days.

The light yellow TSA side for total plate count can be evaluated after 1-2 days.

The Rose Bengal-side affords 3-4 days

If required by legal guidelines or internal regulations, it is possible to deviate from the given recommendations for temperatures and times for incubation.

Limit of detection

The lower detection limit of the germ indicators for bacteria and yeasts is about 100 CFU/ml; for moulds about 10 CFU/ml. If no colonies are visible on the culture medium, the respective germ concentration is below the detection limit.

A differentiation of the microorganisms is only allowed by qualified personnel.

Other materials required

General laboratory equipment, incubator, disinfectant.

Evaluation

Bacteria indicators for liquids allow a semi-quantitative determination of the colony-forming units (= CFU) per ml of the liquid to be examined by comparison with the reading scheme.

Each Dip-Slide can be evaluated after 2 days for bacteria, and then after 3-4 days for yeast and mold. The colonies that have grown are counted. Carefully remove the culture medium from the tube, do not touch it with your fingers!

The colony numbers per culture medium are written down and thus documented.

Colony count x 40 Colony count / ml or CFU / ml (always per side!)

Evaluation diagram

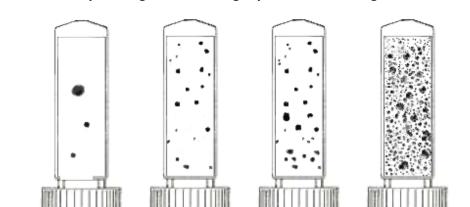
- 1. Bacterial growth on the light yellow nutrient medium. The number of colonies on the agar correlates directly with the number of microorganisms in the sample/surface
- 2. Mold & yeast growth on the light-pink nutrient medium

Especially with molds in liquids, the colony count may vary strongly. The detection of some mold colonies is usually an indicator for strong contamination of the system, because despite mold infestation, spores are not always present.

10⁵



Colony forming units on the light yellow side – total germ count



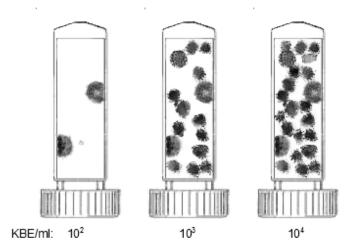
10⁴

Pink side: yeasts

10² 10³ 10⁴

Colony forming units on the pink surface: mold

 10^{3}



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